

Technical Instructions for Spotting Microarrays

PRODUCT

Nexterion™ Slide E is an activated glass slide in the standard size 75.6 mm x 25 mm x 1.0 mm with an epoxy surface coating for efficient covalent and directed binding of molecules, e.g. synthetically fabricated oligonucleotides and/or PCR-products.

ARRAY PRINTING

1. Mix equal amounts of oligonucleotide probe or PCR product and 2X Nexterion Spotting Solution to obtain the recommended final probe concentration (10 to 20 µM for oligonucleotides, 0.1 to 0.5 µM for PCR products).
2. Transfer an appropriate volume of probes to a microtiter plate.
3. Setup the arrayer according to the manufacturers recommendations and print slides.

DNA IMMOBILIZATION

1. Incubate printed microarray slides in humidity chamber consisting of dH₂O for a) 30 min for binding of amine-modified oligonucleotides or b) 30 min for binding of amine-modified PCR products, followed by an exposure to 60°C for 30 min.

When using unmodified probes, incubate the printed microarray slides at 120°C for 30 min in addition to above treatment. For betaine-containing spotting solutions (i.e. 1.5 M betaine in 3X SSC), the microarrays should be incubated at 60° to 120° C for at least 60 min for efficient immobilization of oligonucleotides or PCR-Products.

WASHING

1.
 - a. Rinse 1 x 5 min in 0.1% Triton-X100 at room temperature
 - b. Rinse 2 x 2 min in 1 mM HCl solution at room temperature
 - c. Rinse 1 x 10 min in 100 mM KCl solution at room temperature
 - d. Rinse 1 x 1 min in dH₂O at room temperature
 - e. Proceed to Blocking immediately.

Notes: a) If you have spotted PCR probes, an additional denaturing step is essential after KCl rinse. Boil slides 1 x 3 min in dH₂O at 95 - 100°C.
b) The volume of washing solution should be at least 250 ml for 5 Nexterion Slide E.

BLOCKING

1. Block the slides with Nexterion Blocking Solution or alternatively with 50 mM ethanolamine, 0.1% SDS (add freshly before use) in 0.1 M Tris, pH 9.0 as follows:
 - a. Incubate slides 1 x 15 min in 1X Nexterion Blocking Solution at 50°C and
 - b. Rinse 1 x 1 min in dH₂O at room temperature.
2. Dry the Nexterion Slide E in an oil-free air or nitrogen stream or centrifuge (2 min at 150 to 200x g) to avoid any water stains on the slide surface.

HYBRIDIZATION

1. Re-suspend or dilute the labeled target in Nexterion Hybridization Buffer (alternatively 3-5x SSC and 0.1% SDS) to get at least 90% (v/v) in the final hybridization solution (mixture ratio sample:buffer 1:9).

2. Denature the suspended target by heating at 95°C for 3 min and apply the appropriate volume onto the array surface of a blocked slide.

POST-HYBRIDIZATION WASHING

1. Wash 1 x 10 min in 2X SSC and 0.2% SDS at room temperature.
2. Wash 1 x 10 min in 2X SSC at room temperature.
3. Wash 1 x 10 min in 0.2X SSC at room temperature.
4. Dry the array in an oil free air or nitrogen stream or by centrifugation at 150 to 200x g for 2 min to avoid water stains on the slide surface.

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Complete Protocol

Please find the complete protocol at:

www.us.schott.com/nexterion

(for customers in USA)

www.schott.com/nexterion

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